

Evaluation of methods for extraction of DNA from *Encephalitozoon cuniculi*

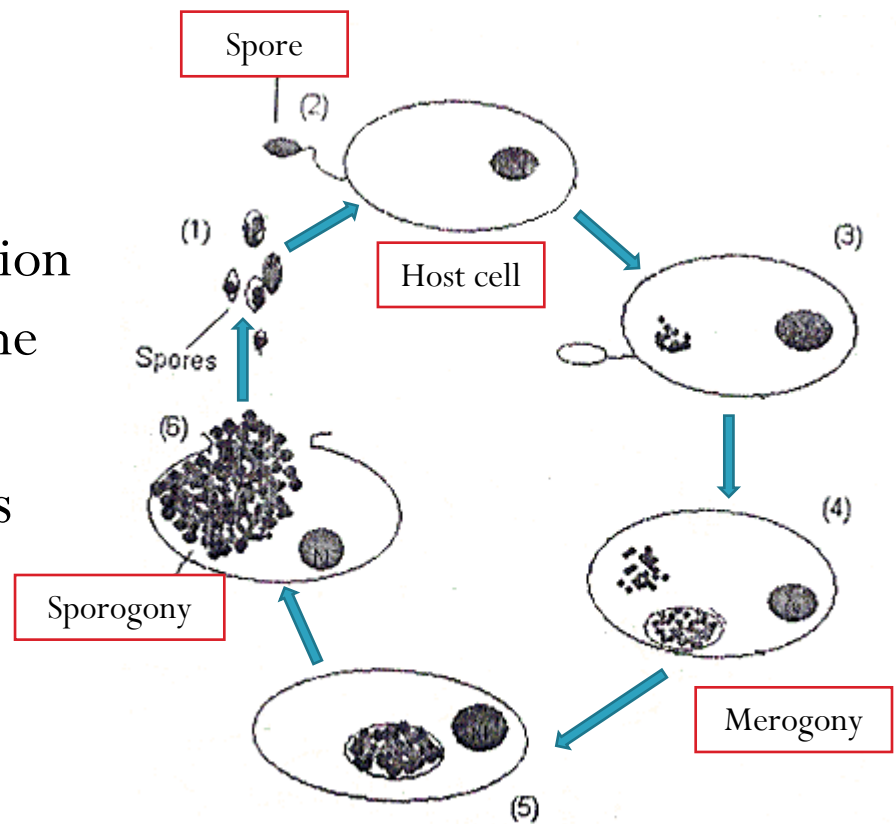
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Encephalitozoonosis

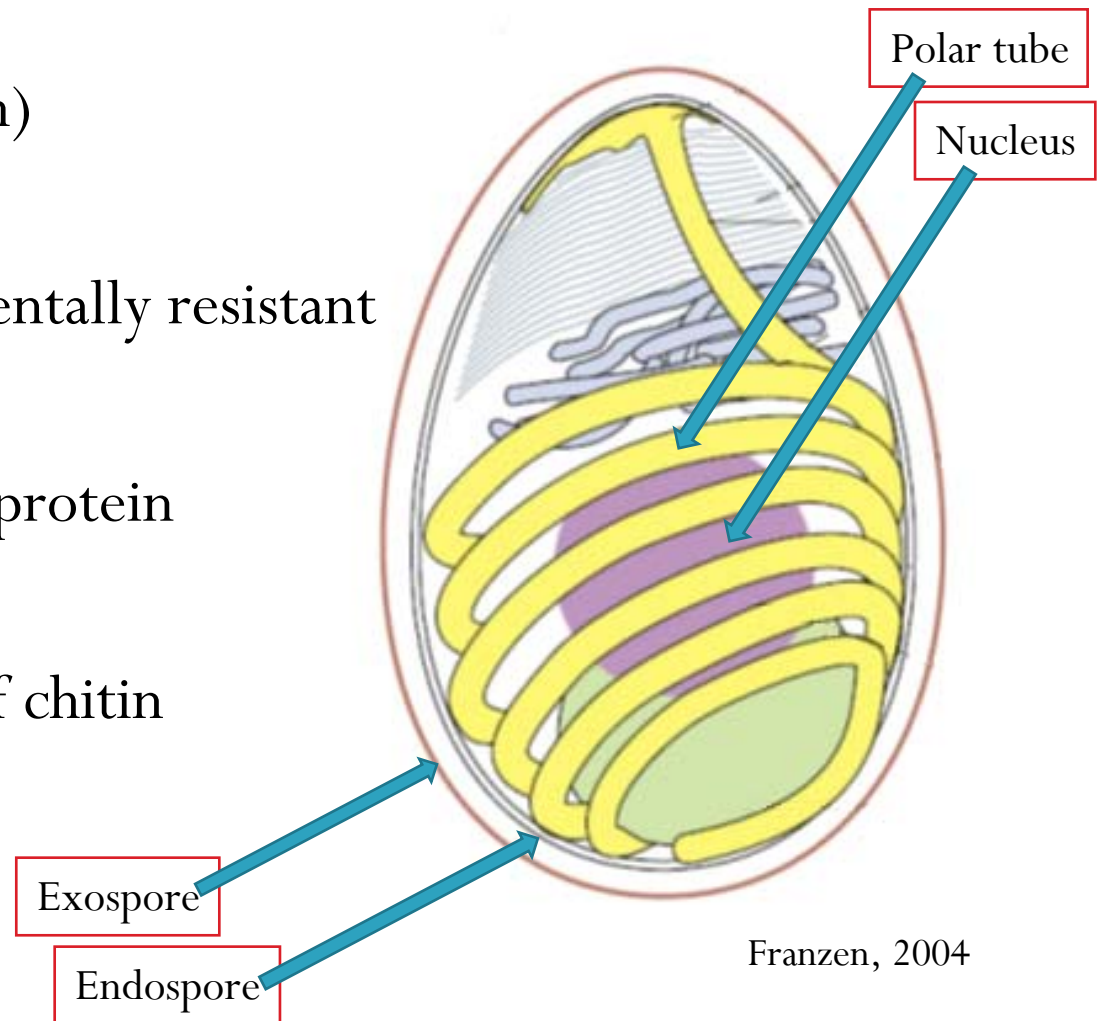


- Disease caused by *Encephalitozoon cuniculi*
 - Fungi (Microsporidia)
 - Unique mechanism of invasion
 - Infective spores shed in urine
- Causes subclinical infections and severe disease
- Zoonotic threat to immunocompromised



Structure of *E. cuniculi* Spores

- Very small (1.5-3 μm)
- Spores are environmentally resistant
- Exospore consists of protein
- Endospore consists of chitin



Franzen, 2004

Diagnostic Techniques

Diagnostic Technique	Limitations
Clinical Presentation	<ul style="list-style-type: none">- Non-specific signs- Differential diagnosis
Serology	<ul style="list-style-type: none">- Indicative of exposure only
Cell Culture	<ul style="list-style-type: none">- Time consuming
Microscopy	<ul style="list-style-type: none">- Spores may be excreted in small numbers- Small size of spores makes identification difficult

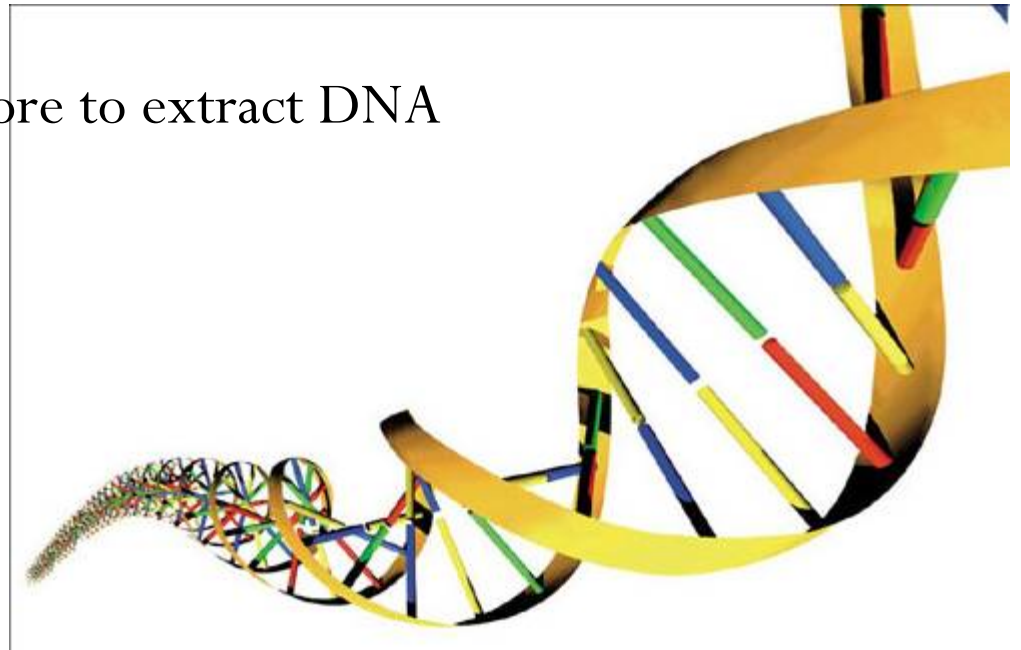
PCR



- Potential to be a rapid and highly sensitive method of detection
- Under-developed in veterinary medicine
- Concerns regarding the sensitivity
 - Moderate sensitivity reported in literature
 - Intermittent shedding of spores or vs. inadequate assay design

DNA Extraction

- No standardized method of DNA extraction
- Sensitivity of PCR depends on the quality and quantity of DNA extraction
- Must first disrupt thick spore to extract DNA
- Enzymatical disruption
- Mechanical disruption



Objectives and Hypothesis

- Objective: To assess the efficacy of different DNA extraction methods for preparation of DNA for PCR
- Hypotheses:
 - 1) A combination of enzymatical and mechanical spore disruption following by DNA extraction with commercially available DNA extraction kits will provide sufficient quantity of DNA to be used for PCR
 - 2) There will be variation in the quantity of DNA extracted amongst the DNA extraction methods

Materials and Methods

- Preparation of spores:
 - Using a reference strain of *E. cuniculi* grown in cell culture
 - Purified and counted (100,000 spores per sample)
 - Enzymatical, mechanical or combination disruption of spores
- DNA extraction:
 - Using one of six commercial kits
 - Samples performed in triplicate
 - Quality and quantity measured using spectrophotometry

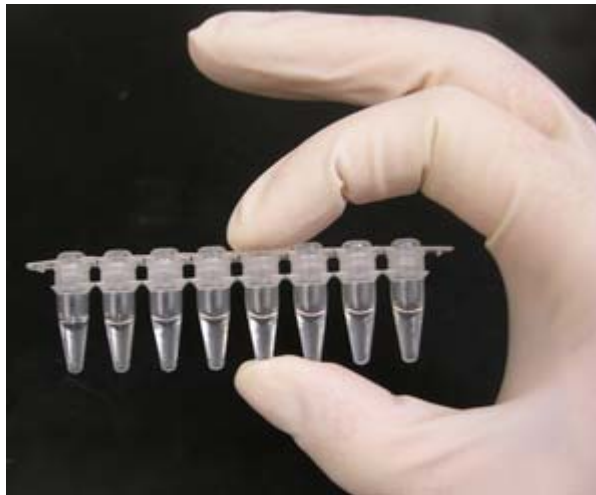
Extraction Kits Tested

- PrepGEM™ (Zygem)
- DNAzol® (Invitrogen)
- Maxwell® 16 Tissue DNA Purification kit (Promega)
- High Pure PCR Template Preparation Kit (Roche)
- DNeasy Plant Mini Kit (Qiagen)
- QIAamp® DNA Stool Mini Kit (Qiagen)



Materials and Methods

- PCR analysis:
 - Using *E. cuniculi* specific primers
 - 50 ng of DNA per reaction
 - Quality of DNA assessed using agarose gel electrophoresis



Results – Experiment #1

- Mean DNA concentration (ng/μl) ± SD:

	DNeasy	Maxwell	QIAamp	High Pure	DNAzol	PrepGE M
No treatment	2.18 (±0.35)	2.85 (±0.03)	7.56 (±1.58)	9.87 (±4.35)	1.67 (±2.69)	8.27 (±10.31)
Mechanical	1.94 (±0.50)	3.67 (±1.57)	4.66 (±1.13)	12.81 (±5.83)	1.47 (±5.92)	5.00 (±4.11)
Enzymatical	3.18 (±0.68)	3.79 (±0.94)	5.59 (±1.21)	11.55 (±0.98)	1.66 (±1.40)	88.44 (±4.11)
Combination	2.91 (±2.12)	4.10 (±0.94)	4.42 (±0.71)	12.68 (±1.38)	1.41 (±2.50)	91.74 (±3.63)

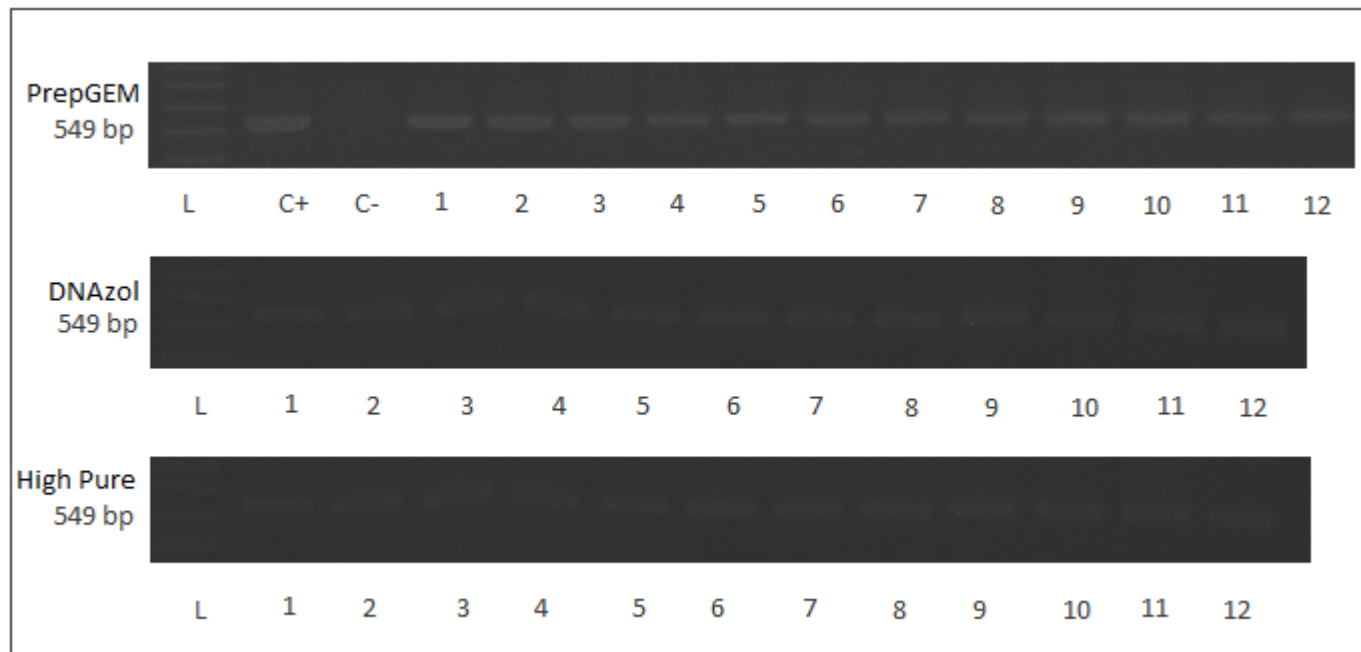
Results – Experiment #2

- Mean DNA concentration (ng/μl) ± SD:

	High Pure	DNAzol	PrepGEM
No treatment	10.25 (±2.93)	3.47 (±1.21)	4.16 (±1.49)
Mechanical	14.70 (±3.71)	0.79 (±0.13)	12.41 (±1.93)
Enzymatical	8.35 (±1.96)	1.69 (±0.82)	148.36 (±2.56)
Combination	23.60 (±20.08)	1.25 (±0.78)	161.63 (±0.52)

Results – Experiment #2

- Agarose gel electrophoresis:



L= ladder, C+= positive control, C-= negative control, 1-3= no treatment, 4-6= mechanical, 7-9= enzymatical, 10-12= combination

Overall Comparison

	Extraction Efficiency	Quality (OD ratio)	Processing Time (m:s)	Difficulty of Procedure	Cost Per Sample (\$)
DNeasy	0.05	2.80	42:10	Difficult	3.56
Maxwell	0.07	1.10	45:40	Easy	5.38
QIAamp	0.11	2.11	34:20	Difficult	3.54
High Pure	0.24 (0.17)	1.39 (1.54)	17:10	difficult	3.19
DNAzol	0.21 (0.02)	3.47 (1.20)	13:10	Moderate	2.45
PrepGEM	1 (1)	0.81 (0.81)	12:30	Easy	1.40

Extraction Efficiency = average yield of kit / average yield of best kit

A_{260}/A_{280} of pure DNA = 1.8-2.2

Conclusions

- A combination of enzymatical and mechanical disruption followed by DNA extraction using PrepGEM produced the highest quantity of DNA
- There is considerable variation in the quantity of DNA extracted amongst the DNA extraction methods tested
- Current methods for extraction of DNA from *E. cuniculi* need to be further evaluated

Future Studies

- Development of a Real time PCR assay
- Detection threshold study

Thank-you